



Revised Structure and Synthesis of Flavolipin

Masao Shiozaki^{a*}, Noriko Deguchi^a, Takashi Mochizuki^a, Takanori Wakabayashi^a,
Tomio Ishikawa^b, Hideyuki Haruyama^b, Yohko Kawai^c and Masahiro Nishijima^c

*a) Exploratory Chemistry Research Laboratories, Sankyo Company, Limited
Hiromachi 1-2-58, Shinagawa-ku, Tokyo 140-8710, Japan*

*b) Analytical and Metabolic Research Laboratories, Sankyo Company, Limited
Hiromachi 1-2-58, Shinagawa-ku, Tokyo 140-8710, Japan*

*c) Department of Biochemistry and Cell Biology, National Institute of Infectious Diseases
Toyama 1-23-1, Shinjuku-ku, Tokyo 162-8640, Japan*

Received 22 June 1998; accepted 24 July 1998

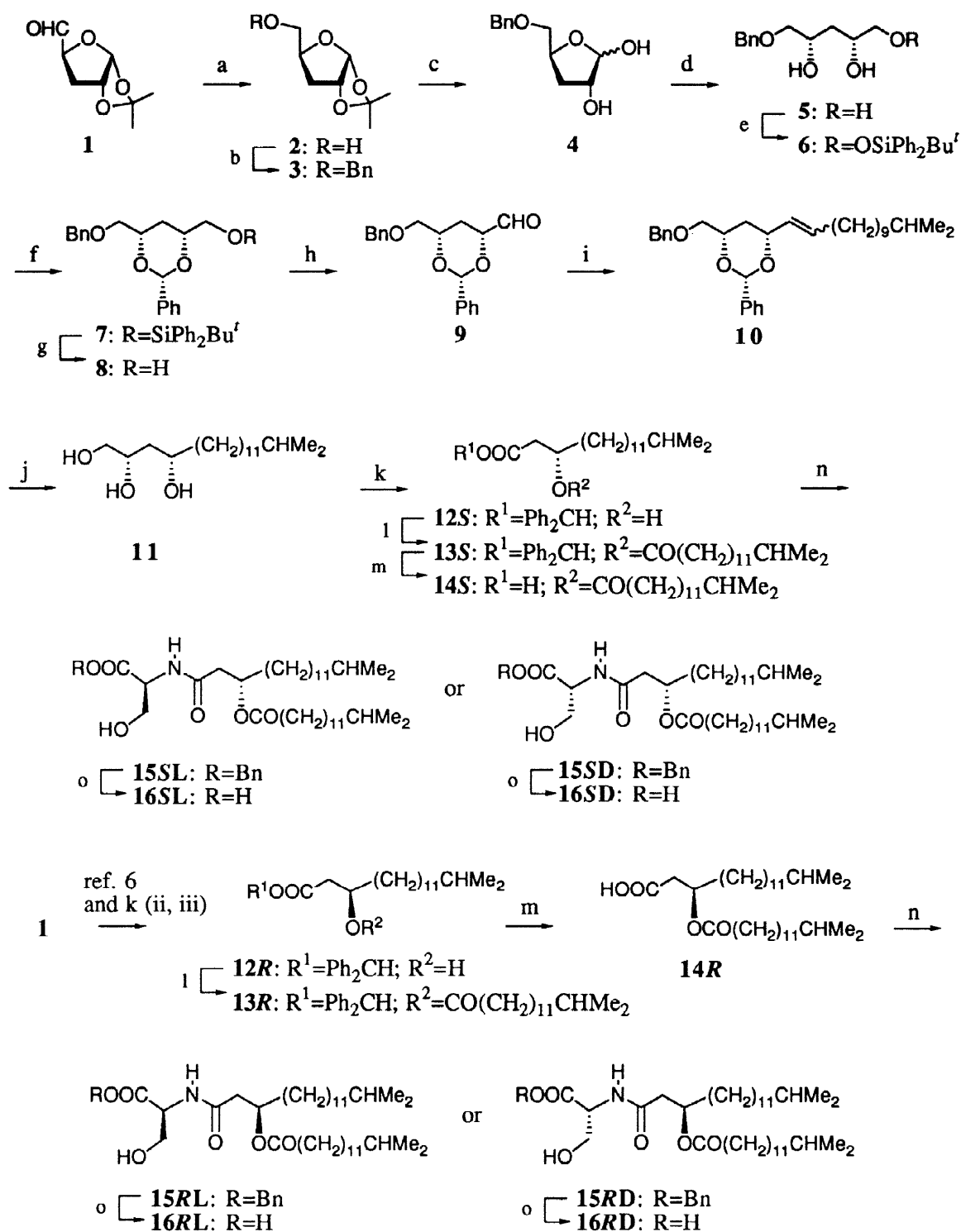
Abstract: The proposed structure of natural flavolipin was revised as *N*-[*N*-[(3*R*)-15-methyl-3-(13-methyltetradecanoyloxy)hexadecanoyl]glycyl]-*L*-serine as a result of a synthetic study and biological activity tests, and its isomers were synthesized in a stereocontrolled manner. © 1998 Elsevier Science Ltd. All rights reserved.

Keywords: stereoisomerism; lipids; peptides; biologically active compounds

Introduction

A serine-containing lipid, flavolipin [1], which was isolated by Kawai *et al.* from an opportunistic pathogen, *Flavobacterium meningosepticum*, exhibits definite hemagglutination activity [1] and strongly stimulates macrophages to generate immunoregulatory substances [2,3]. However, in mice, flavolipin exhibits none of the lethal toxicity [2] that is exhibited by lipopolysaccharide. This fact suggests that flavolipin is a nontoxic immunoactivator [4]. Therefore, we tried to synthesize the proposed flavolipin (**16**) to determine the configuration of the natural flavolipin and to investigate the biological activities of its stereoisomers. All four stereoisomers (**16SL**, **16SD**, **16RL** and **16RD**) of the proposed flavolipin were synthesized in a stereocontrolled manner from D-glucose. However, none of them was identical with the reported natural product in the analytical data [5]. As a result, our synthetic study negated the proposed structure (**16**) [6]. Judging from the FAB MS and ¹H NMR analyses of natural flavolipin, the proposed flavolipin is lacking in a glycine moiety. Moreover, the natural flavolipin yielded negative results in the ninhydrin test; characteristic of amino acids. Therefore, we anticipated the structure of flavolipin as *N*-[*N*-[15-methyl-3-(13-methyltetradecanoyloxy)hexadecanoyl]seryl]glycine (**18**) or *N*-[*N*-[15-methyl-3-(13-methyltetradecanoyloxy)hexadecanoyl]glycyl]serine (**22**) rather than such amino acids as **23**, **24** and **25** in

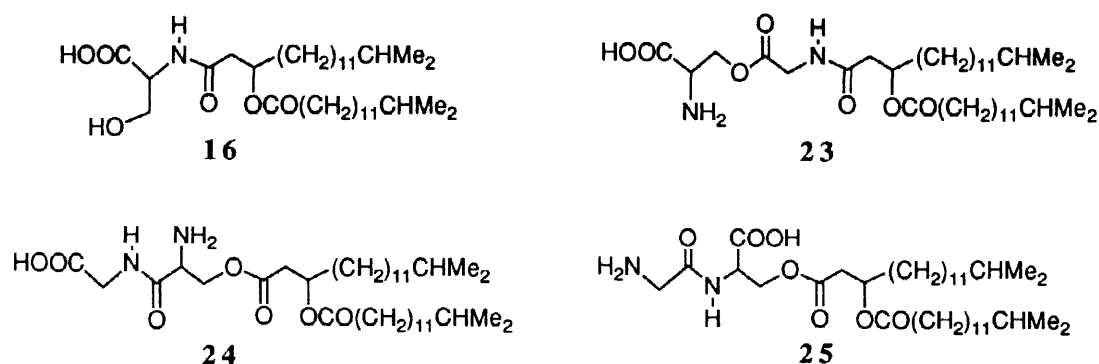
Scheme 1



Reagents and conditions: a) NaBH₄, EtOH, 92%; b) BnBr, NaH, DMF, 80%; c) dioxane-H₂O-1 M HCl (20:1:1), 60%; d) NaBH₄, EtOH, 94%; e) TBDPSCl, Et₃N, 79%; f) PhCH(OMe)₂, PPTS, DMF, 78%; g) Bu₄NF, THF, 98%; h) (COCl)₂, DMSO, Et₃N, CH₂Cl₂, 93%; i) Me₂CH(CH₂)₉CH=PPh₃, THF, 63%; j) H₂, 5% Pd/C, THF; then H₂, Pd(OH)₂/C, MeOH, 65%; k) (i) NaIO₄, dioxane-H₂O (4:1); (ii) *m*-CPBA, CHCl₃; (iii) Ph₂CN₂, EtOAc, 3 steps 83%; l) Me₂CH(CH₂)₁₁COOH, DCC, DMAP, CH₂Cl₂; m) H₂, Pd(OH)₂/C, EtOH, 52% (2 steps); n) L- or D-serine benzyl ester, DCC, DMAP; o) H₂, Pd/C, 57-77% (2 steps).

Figure 1. After synthesizing each of the four stereoisomers of **18** and **22**, and comparing the physical data and biological activity of natural flavolipin with those of synthetic compounds, we revised the proposed structure of natural flavolipin to *N*-[*N*-[(3*R*)-15-methyl-3-(13-methyltetradecanoyloxy)hexadecanoyl]glycyl]-*L*-serine (**22RL**). However, this compound was already reported as WB-3559 D [7] isolated from *Flavobacterium* sp. No. 3559 and topostin D654 [8] isolated from *Flexibacter topostinus* sp. nov. Therefore, it becomes clear that flavolipin is the same compound as both WB-3559 D and topostin D654. In this paper, we describe the synthesis of flavolipin (**22RL**) and all its stereoisomers.

Figure 1



Results and Discussion

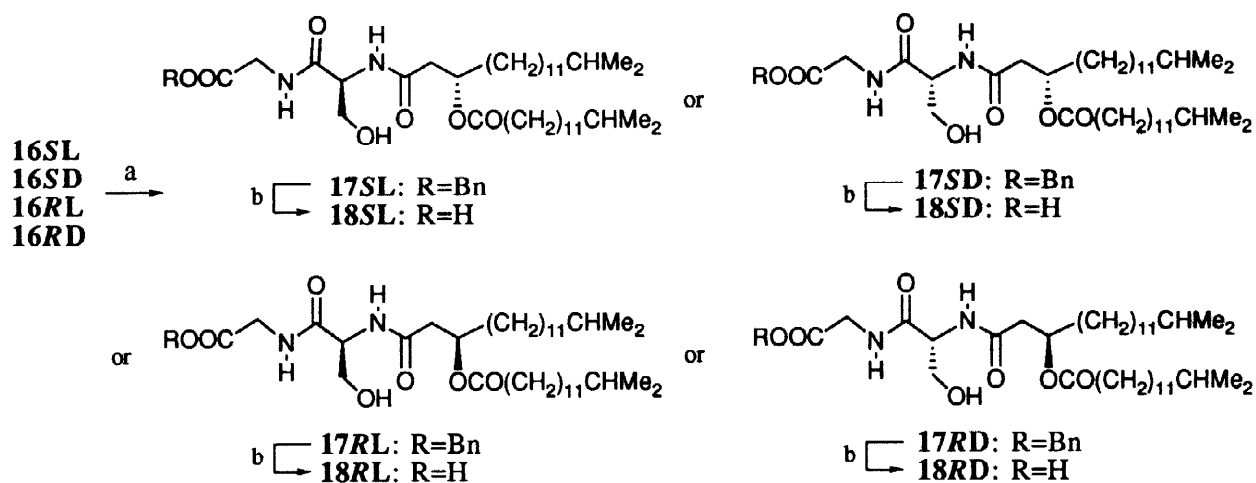
First, we tried to synthesize the proposed flavolipin **16** [2] via (3*S*)- and (3*R*)-15-methyl-3-(13-methyltetradecanoyloxy)hexadecanoic acids (**14S** and **14R**), both of which were obtained from the common aldehyde **1** [9,10,11].

Compound **14S** was synthesized as follows. Reduction of **1** with NaBH₄, followed by benzylation of the resultant alcohol **2** with benzyl bromide and NaH yielded benzyl ether **3**. Treatment of **3** with dioxane-H₂O-1 M aqueous HCl (20:1:1) gave hemiacetal **4**, which was further converted to triol **5** by NaBH₄ reduction. The primary alcohol of **5** was protected as *t*-butyldiphenylsilyl ether **6**, and then the remaining diol was protected as a benzylidene group by treatment with PhCH(OMe)₂ and pyridinium *p*-toluenesulfonate (PPTS) to give **7** as a single isomer. Deprotection of the silyl ether of **7** with tetrabutylammonium fluoride (TBAF), Swern oxidation of the alcohol **8**, and Wittig reaction of the aldehyde **9** with 11-methyldodecyltriphenylphosphorane gave **10**. Compound **10** was hydrogenated over Pd on carbon, and then Pd(OH)₂ on carbon (wet, Degussa type) to give triol **11**. The vicinal diol part of **11** was oxidatively cleaved by NaIO₄, and resultant aldehyde was subjected to *m*-chloroperoxybenzoic acid oxidation. Subsequent esterification of the obtained carboxylic acid by Ph₂CN₂ gave ester **12S**. Reaction of **12S** with 13-methyltetradecanoic acid [7,12] using 1,3-dicyclohexylcarbodiimide (DCC) as dehydrating agent gave ester **13S**. Subsequent hydrogenolysis over Pd(OH)₂ on carbon as a catalyst furnished acid **14S**.

Compound **14R** was obtained from aldehyde **1** according to the reported method [7,12] via compounds **12R** and **13R**.

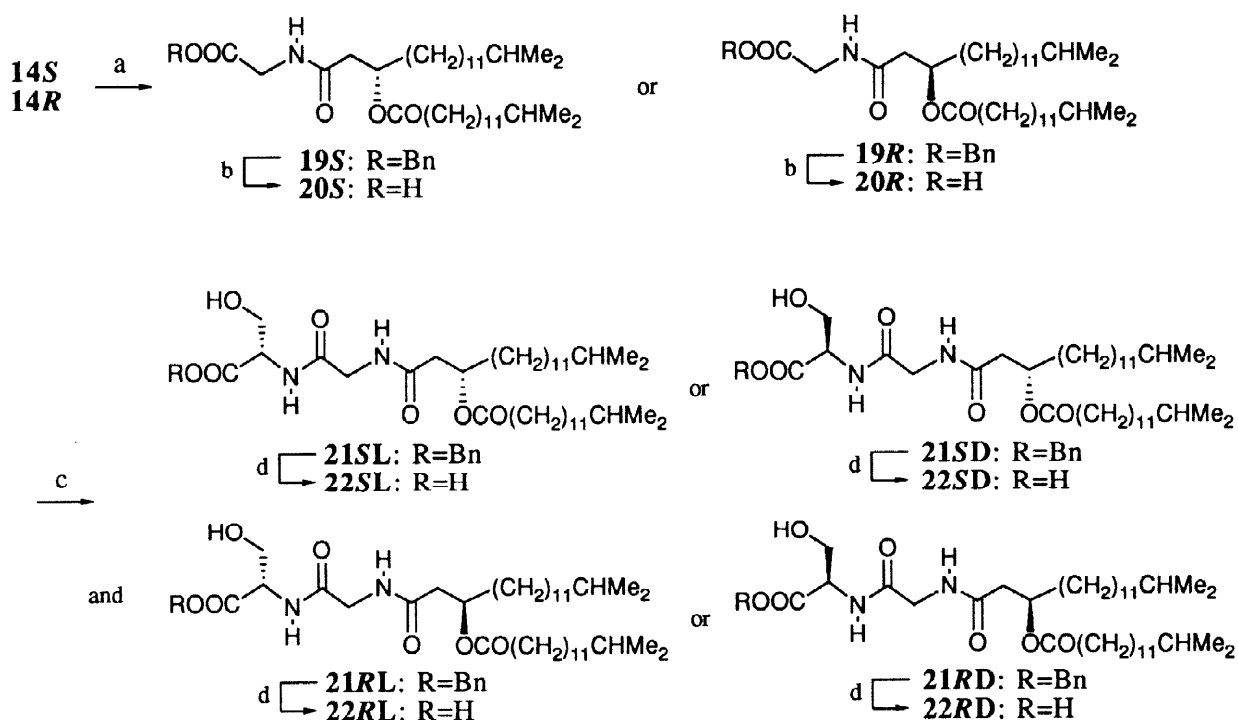
Reaction of **14S** and **14R** with *L*- or *D*-serine benzyl ester using DCC as a dehydrating agent gave benzyl esters **15SL**, **15SD**, **15RL** and **15RD**, respectively. Hydrogenolysis of these four esters produced **16SL** ($[\alpha]_{\text{D}}^{24} = +12.8^\circ$ (c 0.13, CHCl₃)), **16SD** ($[\alpha]_{\text{D}}^{24} = -13.7^\circ$ (c 0.18, CHCl₃)), **16RL** ($[\alpha]_{\text{D}}^{24} = +13.3^\circ$ (c 0.14, CHCl₃)), and **16RD** ($[\alpha]_{\text{D}}^{24} = -14.4^\circ$ (c 0.18, CHCl₃)), respectively. However, none of these four compounds, thus synthesized in a stereocontrolled manner, was identical with the natural flavolipin [5]. Judging

Scheme 2



Reagents and conditions: a) glycine benzyl ester, DCC, CH₂Cl₂, 24 °C, 16 h, 70-91%; b) H₂, Pd/C, EtOAc, 2.5 h, 71-85%.

Scheme 3



Reagents and conditions: a) (COCl)₂, CH₂Cl₂, 24 °C, 1 h, then glycine benzyl ester hydrochloride, Et₃N, CH₂Cl₂, 24 °C, 1 h, 57-69%; b) H₂, Pd/C, EtOAc, 2 h, 94-98%; c) L or D-serine benzyl ester, DCC, CH₂Cl₂, 24 °C, 2 h, 70-82%; d) H₂, Pd/C, EtOAc, 2.5 h, 50-73%.

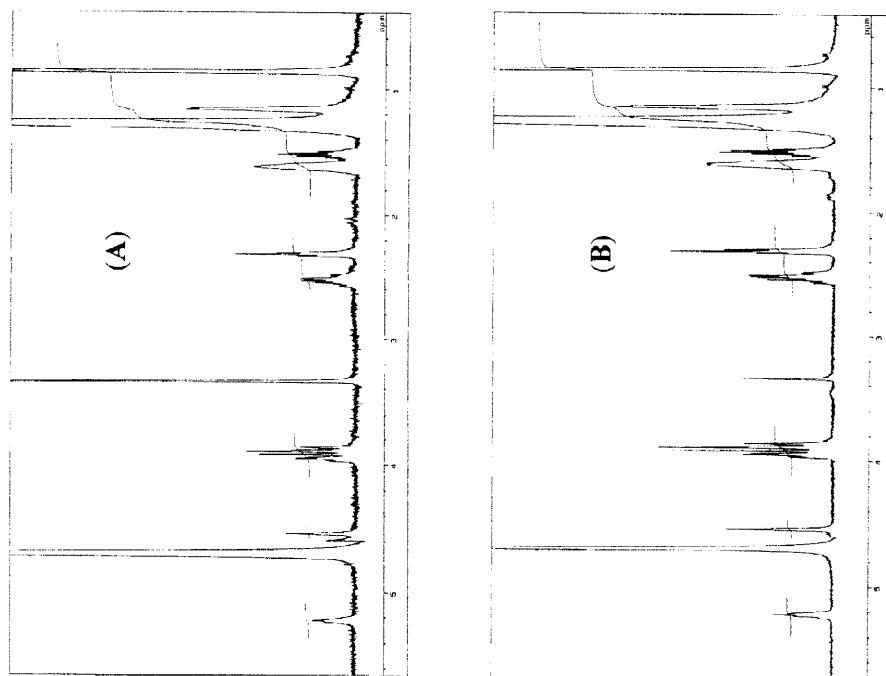


Figure 2. 400 MHz ¹H NMR (CD₃OD) of Natural Flavolipin (A) and Synthetic Flavolipin (B).

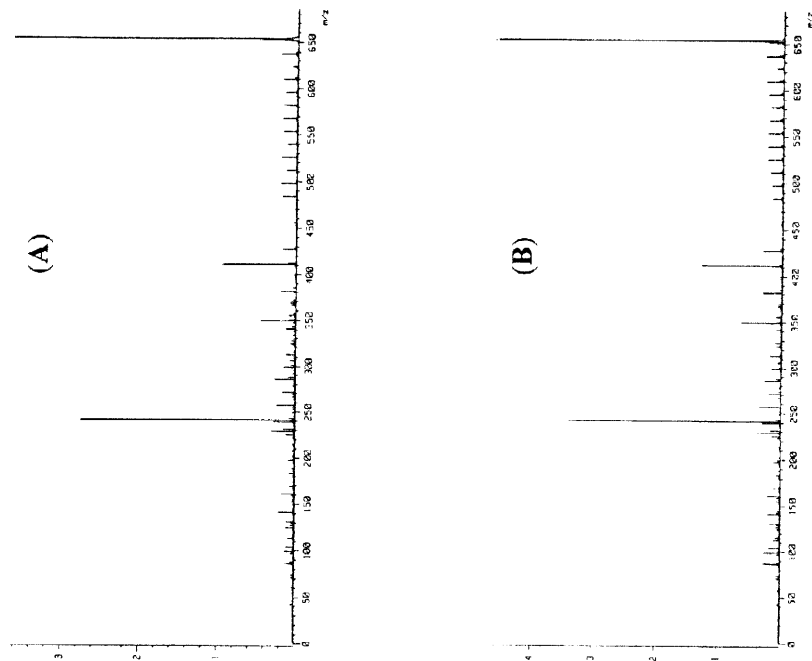


Figure 3. FAB negative MS of Natural Flavolipin (A) and Synthetic Flavolipin (B).

from the physical data of natural flavolipin, it was apparent that the proposed structure of flavolipin was lacking in a glycine part.

Therefore, next, we synthesized the acyl-serylglycines **18**. Treatment of **16SL**, **16SD**, **16RL**, and **16RD** with glycine benzyl ester using DCC as a condensing reagent gave the corresponding amide benzyl esters **17SL**, **17SD**, **17RL**, and **17RD**, respectively. Hydrogenolytic deprotection of each benzyl ester gave the corresponding acids **18SL** ($[\alpha]_{\text{D}}^{24} -8.0^\circ$ (c 0.23, CHCl_3)), **18SD** ($[\alpha]_{\text{D}}^{24} +7.1^\circ$ (c 0.63, CHCl_3)), **18RL** ($[\alpha]_{\text{D}}^{24} -7.8^\circ$ (c 0.50, CHCl_3)), and **18RD** ($[\alpha]_{\text{D}}^{24} +7.6^\circ$ (c 0.75, CHCl_3)). Disappointingly, none of these compounds was identical to the natural flavolipin.

Further, we tried to synthesize acyl-glycylserines **22**. Compounds **14S** and **14R** were converted with oxalyl chloride to their corresponding acid chlorides, which were treated with glycine benzyl ester hydrochloride and Et_3N to give the corresponding amides **19S** and **19R**. Then, hydrogenolytic deprotection of each benzyl ester of the amides gave the corresponding acids **20S** [13] and **20R** [8,14], which were treated with L- and D-serine benzyl esters using DCC as a condensing reagent to give the corresponding amides **21SL**, **21SD**, **21RL**, and **21RD**, respectively. Subsequent hydrogenolytic deprotection of each benzyl ester gave the corresponding acids **22SL** ($[\alpha]_{\text{D}}^{24} +25.6^\circ$ (c 0.38, CHCl_3)), **22SD** ($[\alpha]_{\text{D}}^{24} -18.8^\circ$ (c 0.52, CHCl_3)), **22RL** ($[\alpha]_{\text{D}}^{24} +18.9^\circ$ (c 0.39, CHCl_3)) [15], and **22RD** ($[\alpha]_{\text{D}}^{24} -26.2^\circ$ (c 0.58, CHCl_3)).

Fortunately, the ^1H NMR and FAB MS spectra of compounds **22SD** and **22RL** were identical to natural flavolipin as shown in Figure 2 and Figure 3.

The macrophage stimulation activity of **22RL** was almost the same as that of natural flavolipin; and **22SD**, however, was practically inactive. Therefore, from this synthetic study we clearly determined the structure of natural flavolipin to be *N*-[*N*-[(3*R*)-15-methyl-3-(13-methyltetradecanoyloxy)hexadecanoyl]glycyl]-*L*-serine [7,8].

Conclusion

Thus we could synthesize all eight isomers of flavolipin in a stereocontrolled manner and determine the correct structure of flavolipin. We are now investigating the biological activities of all the isomeric compounds of **18** and **22**, and the results will be reported in due course.

Experimental

Melting points were determined on a Yanagimoto micro melting point apparatus and were uncorrected. ^1H NMR (270 and 400 MHz) spectra were recorded with JEOL JNM-270 and JNM-GSX 400 spectrometers using TMS as an internal standard. IR absorption spectra were determined with a IR A-2 spectrophotometer, and mass spectra were obtained with a JMS-700 mass spectrometer. Optical rotations were obtained by the use of a Perkin-Elmer 241 polarimeter. Elemental analyses were performed by the Institute of Science and Technology, Inc. Separation of the compounds by column chromatography was done with silica gel 60 (230-400 mesh ASTM, E. Merck) under a slightly elevated pressure (1.2-1.5 atm) for easy elution, and the quantity of silica gel used was 50-100 times the weight charged on the column. Preparative TLC was performed on silica gel plates (Merck, Silica Gel 60 F245). Detection involved spraying the chromatogram with a solution of 17% H_2SO_4 in water (w/w), containing ammonium molybdate (2.3%) and ceric sulfate (0.9%) (Hanessian dip), and heating the plate for several minutes at ca 180°C . Tetrahydrofuran (THF) was distilled from LiAlH_4 and used immediately. CH_2Cl_2 was dried by being passed through an ICN Alumina B-Super I. *N,N*-Dimethylformamide (DMF) and pyridine were dried by storage over 4\AA molecular sieves. MeCN was dried by storage over 3\AA molecular sieves.

3-Deoxy-1,2-O-isopropylidene- α -D-ribose (2). To a solution of **1** [9,10,11] (4.80 g, 27.9 mmol) in 99.5% EtOH (200 ml) was added NaBH₄ (1.06 g, 28.0 mmol) at 0–5 °C. After stirring for 30 min, the reaction mixture was diluted with CHCl₃, washed with H₂O, and brine, dried over MgSO₄, and filtered. The filtrate was concentrated *in vacuo* to give **2** (4.45 g, 92%). IR ν_{\max} (KBr) 3482, 3382 cm⁻¹. 270 MHz ¹H NMR (CDCl₃) δ 1.33 (3H, s), 1.52 (3H, s), 1.77–1.90 (2H, m, containing OH), 2.01 (1H, dd, *J*=4.6, 13.5 Hz), 3.57 (1H, ddd, *J*=4.5, 7.1, 11.6 Hz), 3.90 (1H, td, *J*=3.2, 12.4 Hz), 4.35 (1H, m), 4.76 (1H, t, *J*=4.2 Hz), 5.83 (1H, d, *J*=3.7 Hz). EI MS *m/z* 175 (M+1). Anal. Calcd. for C₈H₁₄O₄·0.1H₂O (174.2+1.8): C, 54.55; H, 8.13. Found: C, 54.70; H, 8.20.

5-O-Benzyl-3-deoxy-1,2-O-isopropylidene- α -D-ribose (3). A solution of **2** (7.24 g, 41.6 mol) in DMF (80 ml) was added dropwise to a suspension of NaH (1.05 g, 43.8 mmol) at 0–5 °C. After stirring for 30 min at 24 °C, benzyl bromide (5.2 ml, 43.7 mmol) was added to this mixture, which was stirred for 16 h at 24 °C. The reaction mixture was diluted with EtOAc, washed with H₂O and brine, dried over MgSO₄, and filtered. The filtrate was concentrated *in vacuo* to give a residue, which was chromatographed on a silica gel column. Elution with cyclohexane–EtOAc (9:1) gave **3** (8.81 g, 80%). IR ν_{\max} (CHCl₃) 2992, 2937 cm⁻¹. 270 MHz ¹H NMR (CDCl₃) δ 1.32 (3H, s), 1.51 (3H, s), 1.77 (1H, ddd, *J*=4.9, 10.9, 13.4 Hz), 2.06 (1H, dd, *J*=4.5, 13.4 Hz), 3.55 (1H, dd, *J*=4.9, 10.7 Hz), 3.65 (1H, dd, *J*=3.6, 10.7 Hz), 4.40 (1H, m), 4.59 (2H, s), 4.73 (1H, t, *J*=4.2 Hz), 5.84 (1H, d, *J*=3.6 Hz), 7.26–7.40 (5H, m). EI MS *m/z* 264 (M⁺). Anal. Calcd. for C₁₅H₂₀O₄ (264.3): C, 68.16; H, 7.63. Found: C, 68.15; H, 7.62.

5-O-Benzyl-3-deoxy-D-ribose (4). A solution of **3** (8.94 g, 33.8 mmol) in 1,4-dioxane (200 ml), H₂O (10 ml) and aqueous 1 M HCl (10 ml) was stirred for 3 h at 70 °C. The reaction mixture was neutralized with 1 M aqueous NaOH (10 ml), concentrated *in vacuo*, diluted with CHCl₃, washed with H₂O, dried over MgSO₄, and filtered. The filtrate was concentrated *in vacuo* to give a residue, which was chromatographed on a silica gel column. Elution with cyclohexane–EtOAc (3:1) gave **4** (4.55 g, 60%). IR ν_{\max} (CHCl₃) 3417, 3086, 1717 cm⁻¹. 270 MHz ¹H NMR (CDCl₃) δ 1.87–2.09 (1.35H, m), 2.25 (0.65H, ddd, *J*=5.1, 8.3, 13.5 Hz), 2.54 (0.35H, d, *J*=6.6 Hz, OH), 3.40–3.59 (1.35H, m), 3.68 (0.65H, dd, *J*=2.7, 10.1 Hz), 3.77 (0.65H, d, *J*=7.9 Hz, OH), 4.22 (0.65H, t, *J*=4.8 Hz, changed to a doublet on addition of D₂O), 4.30 (0.35H, m, changed to dd, *J*=5.2, 11.0 Hz on addition of D₂O), 4.44–4.62 (3H, m), 5.17 (0.65H, d, *J*=8.0 Hz, changed to a singlet on addition of D₂O), 5.38 (0.35H, t, *J*=4.0–5.0 Hz, changed to a doublet, *J*=4.0 Hz, on addition of D₂O), 7.26–7.39 (5H, m). MS *m/z* 225 (M⁺+1), 224 (M⁺). Anal. Calcd. for C₁₂H₁₆O₄ (224.3): C, 64.27; H, 7.19. Found: C, 64.38; H, 7.11.

(2S,4R)-1-Benzyl-2,4,5-trihydroxypentane (5). To a solution of **4** (3.36 g, 15.0 mmol) in 99.5% EtOH (50 ml) was added NaBH₄ (570 mg, 15 mmol) at 0–5 °C. After stirring for 1.5 h at 0–5 °C, the reaction mixture was quenched with 0.1 M aqueous HCl and diluted with EtOAc. The solution was washed with H₂O and brine, dried over MgSO₄, and filtered. The filtrate was concentrated *in vacuo* to give a residue, which was chromatographed on a silica gel column. Elution with CH₂Cl₂–MeOH (20:1) gave **5** (3.18 g, 94%) as an oil. IR ν_{\max} (CHCl₃) 3584, 3474 cm⁻¹. 270 MHz ¹H NMR (CDCl₃) δ 1.59–1.65 (2H, m), 2.30 (1H, bs), 3.03 (1H, bs), 3.35–3.66 (5H, m), 3.96–4.12 (2H, m), 4.56 (2H, s), 7.26–7.36 (5H, m). MS *m/z* 227 (M⁺+1). Anal. Calcd. for C₁₂H₁₈O₄·0.5 H₂O (226.3+9.0): C, 61.25; H, 8.14. Found: C, 61.16; H, 8.15.

(2R,4S)-5-Benzyl-1-(tert-butyl-diphenylsilyloxy)-2,4-dihydroxypentane (6). To a solution of **5** (300 mg, 1.33 mmol) in DMF (5 ml) were added *tert*-butyl-diphenylsilyl chloride (0.38 ml, 1.46 mmol) and Et₃N (0.20 ml, 1.44 mmol) at 24 °C under nitrogen. After stirring for 2 h at 24 °C, the reaction

mixture was diluted with EtOAc. The solution was sequentially washed with 0.1 M HCl, H₂O, sat. NaHCO₃, and brine, dried over MgSO₄, and filtered. The filtrate was concentrated *in vacuo* to give a residue, which was chromatographed on a silica gel column. Elution with hexane-EtOAc (3:1) gave **6** (488 mg, 79%) as an oil. IR $\nu_{\text{max}}(\text{CHCl}_3)$ 3583, 3517 cm⁻¹. 270 MHz ¹H NMR (CDCl₃) δ 1.06 (9H, s), 1.51–1.70 (2H, m), 3.09 (1H, d, *J*=2.5 Hz, OH), 3.22 (1H, d, *J*=1.9 Hz, OH), 3.41, 3.45 (2H, AB-q, *J*=10.4 Hz), 3.55–3.63 (2H, m), 3.97–4.06 (2H, m), 4.55 (2H, s), 7.28–7.47 (11H, m), 7.63–7.67 (4H, m). FAB MS (positive) *m/z* 487 [M+Na]⁺. High Resolution FAB MS, Calcd. for C₂₈H₃₆O₄SiNa: 487.2280; Found: 487.2254.

(2R,4S)-2,4-[(R)-Benzylidenedioxy]-5-benzyloxy-1-(tert-butylidiphenylsilyloxy)pentane (7). To a solution of **6** (480 mg, 1.03 mmol) in DMF (10 ml) were added PhCH(OMe)₂ (5.0 ml) and pyridinium *p*-toluenesulfonate (1.0 g, 3.98 mmol) at 24 °C under nitrogen atmosphere. After stirring for 16 h at 24 °C, the reaction mixture was diluted with EtOAc. The solution was washed with sat. NaHCO₃ and brine, dried over MgSO₄, and filtered. The filtrate was concentrated *in vacuo* to give a residue, which was chromatographed on a silica gel column. Elution with hexane-EtOAc (20:1) gave **7** (448 mg, 78%) as an oil. IR $\nu_{\text{max}}(\text{CHCl}_3)$ 2961, 2932, 2861 cm⁻¹. 270 MHz ¹H NMR (CDCl₃) δ 1.06 (9H, s), 1.53 (1H, m), 3.09 (1H, d, *J*=2.5 Hz, OH), 3.22 (1H, d, *J*=1.9 Hz, OH), 3.41, 3.45 (2H, AB-q, *J*=10.4 Hz), 3.55–3.63 (2H, m), 3.97–4.06 (2H, m), 4.55 (2H, s), 7.28–7.47 (16H, m), 7.63–7.67 (4H, m). EI MS *m/z* 552 (M⁺+1), 551 (M⁺). High Resolution MS, Calcd. for C₃₅H₄₀O₄Si: 552.2696; Found: 552.2672.

(2S,4R)-2,4-[(S)-Benzylidenedioxy]-1-benzyloxy-5-hydroxypentane (8). To a solution of **7** (440 mg, 0.80 mmol) in THF (5 ml) was added 1 M THF solution of TBAF (1.0 ml) at 24 °C under nitrogen. After stirring for 30 min at 24 °C, the reaction mixture was diluted with EtOAc. The solution was washed with H₂O and brine, dried over MgSO₄, and filtered. The filtrate was concentrated *in vacuo* to give a residue, which was chromatographed on a silica gel column. Elution with hexane-EtOAc (2:1) gave **8** (246 mg, 98%) as a solid. mp 53–55 °C (from EtOAc-hexane). IR $\nu_{\text{max}}(\text{CHCl}_3)$ 3601, 2924, 2869 cm⁻¹. 270 MHz ¹H NMR (CDCl₃) δ 1.57–1.62 (2H, m), 2.00 (1H, broad, OH), 3.54 (1H, dd, *J*=5.0, 10.2 Hz), 3.62–3.78 (3H, m), 4.03 (1H, m), 4.15 (1H, m), 4.58, 4.62 (2H, AB-q, *J*=12.2 Hz), 5.60 (1H, s), 7.28–7.42 (8H, m), 7.48–7.54 (2H, m). MS *m/z* 314 (M⁺). Anal. Calcd. for C₁₉H₂₂O₄ (314.4): C, 72.59; H, 7.05. Found: C, 72.31; H, 7.24.

(2R,4S)-2,4-[(R)-Benzylidenedioxy]-5-benzyloxy-1-benzyloxy-5-hydroxypentanal (9). To a solution of oxalyl chloride (0.075 ml, 0.860 mmol) in CH₂Cl₂ (1 ml) was added dimethyl sulfoxide (0.135 ml, 1.90 mmol) at -78 °C with stirring under nitrogen. After 5 min, a solution of **8** (100 mg, 0.32 mmol) in CH₂Cl₂ (1 ml) was added to this solution at -78 °C, and the stirring was continued for 15 min, and then Et₃N (0.60 ml, 4.30 mmol) was added. After 5 min, the reaction mixture was warmed to room temperature and diluted with CH₂Cl₂. The solution was washed with H₂O and brine, dried over MgSO₄, and filtered. The filtrate was concentrated *in vacuo* to give a residue, which was chromatographed on a silica gel column. Elution with hexane-EtOAc (2:1) gave **9** (93 mg, 93%) as an oil. IR $\nu_{\text{max}}(\text{CHCl}_3)$ 1740 cm⁻¹. 270 MHz ¹H NMR (CDCl₃) δ 1.67 (1H, td, *J*=11.6, 13.6 Hz), 1.95 (1H, td, *J*=2.8, 13.6 Hz), 3.57 (1H, dd, *J*=5.3, 9.9 Hz), 3.69 (1H, dd, *J*=5.3, 9.9 Hz), 4.16 (1H, m), 4.36 (1H, dd, *J*=2.8, 11.6 Hz), 4.60 (2H, s), 5.65 (1H, s), 7.29–7.57 (10H, m), 9.74 (1H, s). FAB MS (negative) *m/z* 311 (M-1)⁻. Anal. Calcd. for C₁₉H₂₀O₄ (312.4): C, 73.06; H, 6.45. Found: C, 73.00; H, 6.66.

(2S,4R)-2,4-[(S)-Benzylidenedioxy]-1-benzyloxy-16-methylheptadec-5(EZ)-ene (10).

To a solution of 11-methyldodecyltriphenylphosphonium chloride (1.60 g, 3.33 mmol) in THF (15 ml) was added a 1.6 M hexane solution of *n*-BuLi (2.00 ml, 3.20 mmol) at -78 °C with stirring under nitrogen. After

stirring for 30 min at $-78\text{ }^{\circ}\text{C}$ and then for 30 min at $24\text{ }^{\circ}\text{C}$, this solution was poured into a solution of **9** (100 mg, 0.33 mmol) in THF (10 ml) at $24\text{ }^{\circ}\text{C}$, and stirring was continued for 1 h. The reaction mixture was concentrated *in vacuo*, diluted with EtOAc, washed with H_2O and brine, dried over MgSO_4 , and filtered. The filtrate was concentrated *in vacuo* to give a residue, which was chromatographed on a silica gel column. Elution with hexane-EtOAc (20:1) gave an *E*:*Z*=5:3 mixture of **10** (97 mg, 63%) as an oil. IR $\nu_{\text{max}}(\text{CHCl}_3)$ 2955, 2928, 2856 cm^{-1} . The mixture was partially separated on a silica gel preparative TLC plate. [*E*-isomer: $R_f=0.44$ (benzene); *Z*-isomer: $R_f=0.36$ (benzene)]. 270 MHz ^1H NMR of *E*-isomer: (CDCl_3) δ 0.86 (6H, d, $J=6.6$ Hz), 1.10–1.37 (17H, m), 1.44–1.70 (2H, m), 2.29–2.47 (2H, m), 3.53 (1H, dd, $J=4.9, 10.0$ Hz), 3.68 (1H, dd, $J=5.8, 10.0$ Hz), 4.13 (1H, m), 4.53 (1H, m), 4.58, 4.62 (2H, AB-q, $J=11.9$ Hz), 5.60 (1H, s), 5.60–5.72 (2H, m), 7.28–7.39 (8H, m), 7.47–7.53 (2H, m). 270 MHz ^1H NMR of *Z*-isomer: (CDCl_3) δ 0.86 (6H, d, $J=6.6$ Hz), 1.13–1.45 (17H, m), 1.45–1.65 (2H, m), 2.07–2.17 (2H, m), 3.53 (1H, dd, $J=4.8, 10.1$ Hz), 3.68 (1H, dd, $J=5.9, 10.1$ Hz), 4.16 (1H, m), 4.59, 4.63 (2H, AB-q, $J=12.4$ Hz), 4.68 (1H, dd, $J=7.4, 13.9$ Hz), 5.44–5.59 (2H, m), 5.62 (1H, s), 7.26–7.39 (8H, m), 7.49–7.54 (2H, m). FAB MS (positive) m/z 501 $[\text{M}+\text{Na}]^+$. High Resolution FAB MS (positive) m/z : Calcd. for $\text{C}_{32}\text{H}_{46}\text{O}_3\text{Na}$: 501.3345; Found: 501.3336.

(2*S*,4*S*)-16-Methyl-1,2,4-trihydroxyheptadecane (11). A solution of the *E*:*Z* mixture of **10** (50 mg, 0.18 mmol) in THF (10 ml) containing 5% Pd on carbon (25 mg) was hydrogenated for 1 h at $20\text{--}25\text{ }^{\circ}\text{C}$ and filtered. The filtrate was concentrated *in vacuo* to give a residue, which was dissolved in MeOH (20 ml). The solution was hydrogenolyzed using $\text{Pd}(\text{OH})_2$ on carbon (wet, Degussa type, 20 mg) for 1 h at room temperature. The reaction mixture was filtered through Celite, and concentrated *in vacuo* to give **11** (27 mg, 65%) as a powder, which was partly recrystallized from EtOAc; mp $70\text{ }^{\circ}\text{C}$. IR $\nu_{\text{max}}(\text{CHCl}_3)$ 3307, 2953, 2920, 2850 cm^{-1} . 270 MHz ^1H NMR: (CDCl_3) δ 0.86 (6H, d, $J=6.6$ Hz), 1.10–1.69 (27H, m), 2.46–2.58 (1H, broad, OH), 3.49 (1H, dd, $J=6.6, 11.3$ Hz), 3.65 (1H, dd, $J=3.3, 11.3$ Hz), 3.87–3.98 (2H, m). FAB MS (positive), m/z 325 $[\text{M}+\text{Na}]^+$.

(3*S*)-Diphenylmethyl 15-methyl-3-hydroxyhexadecanoate (12*S*). (i) A solution of **11** (31 mg, 0.10 mmol) in 1,4-dioxane (2 ml) and a solution of NaIO_4 (111 mg, 0.52 mmol) in H_2O (0.4 ml) were mixed and stirred for 1 h at $24\text{ }^{\circ}\text{C}$. The reaction mixture was diluted with H_2O and extracted with CHCl_3 . The solution was washed with brine, dried over MgSO_4 , and filtered. The filtrate was concentrated *in vacuo* to give 28 mg of (3*S*)-15-methyl-3-hydroxyhexadecanal, which was dissolved in CHCl_3 (5 ml). (ii) To this solution was added *m*-chloroperoxybenzoic acid (28 mg, 0.16 mmol). The solution was warmed at $50\text{ }^{\circ}\text{C}$ for 1 h with stirring in the dark to give (3*S*)-15-methyl-3-hydroxyhexadecanoic acid and concentrated *in vacuo* to give a mixture, which was dissolved in EtOAc (5 ml). (iii) To this solution was added diphenyldiazomethane (78 mg, 0.40 mmol). This solution was warmed at $50\text{ }^{\circ}\text{C}$ for 45 min with stirring and concentrated *in vacuo* to give a residue, which was chromatographed on a silica gel column. Elution with benzene-EtOAc (19:1) gave a benzhydryl ester **12*S*** (40 mg, 86%) as an oil. $[\alpha]_{\text{D}}^{24} +14.5^{\circ}$ (c 0.2, CHCl_3). IR $\nu_{\text{max}}(\text{film})$ 3032 (broad), 2926, 2853, 1736 cm^{-1} . 270 MHz ^1H NMR: (CDCl_3) δ 0.86 (6H, d, $J=7.0$ Hz), 1.14–1.55 (23H, m), 2.58–2.61 (3H, m), 4.01 (1H, m), 6.91 (1H, s), 7.33–7.34 (10H, m). EI MS m/z 452 (M^+). Anal. Calcd. for $\text{C}_{30}\text{H}_{44}\text{O}_3\cdot 0.24\text{H}_2\text{O}$ (457.0): C, 78.85; H, 9.81. Found: C, 78.97; H, 9.61.

(3*R*)-Diphenylmethyl 15-methyl-3-hydroxyhexadecanoate (12*R*). The above mentioned procedures (ii) and (iii) applied to (3*R*)-15-methyl-3-hydroxyhexadecanal (833 mg, 3.08 mmol), which was obtained from compound (**1**) according to the reported method [6], gave **12*R*** (1.261 g) in 90% yield as an oil. $[\alpha]_{\text{D}}^{24} -14.0^{\circ}$ (c 0.2, CHCl_3). EI MS m/z 452 (M^+).

(3S)-Diphenylmethyl 15-methyl-3-(13-methyltetradecanoyloxy)hexadecanoate (13S).

To a solution of **12S** (1.12 g, 2.47 mmol) and 13-methyltetradecanoic acid (0.90 g, 3.71 mmol) in CH_2Cl_2 (25 ml) were added 1,3-dicyclohexylcarbodiimide (765 mg, 3.71 mmol) and 4-dimethylaminopyridine (453 mg, 3.71 mmol). The mixture was stirred for 16 h at room temperature and filtered through Celite. After filtration, the filter cake was washed with a small amount of EtOAc. The combined filtrate was diluted with EtOAc. The solution was washed with aq. NaHCO_3 and brine, dried over MgSO_4 , filtered, concentrated *in vacuo*, and the residual mixture was chromatographed on a silica gel column. Elution with hexane-EtOAc (20:1) gave **13S** (1.52 g, 90%) as a wax. $[\alpha]_{\text{D}}^{24} +3.1^\circ$ (c 0.2, CHCl_3). IR ν_{max} (film) 2926, 2855, 1742 cm^{-1} . 270 MHz ^1H NMR: (CDCl_3) δ 0.86 (12H, d, $J=6.6$ Hz), 1.13–1.55 (44H, m), 2.05–2.15 (2H, m), 2.61–2.70 (2H, m), 5.26 (1H, m), 6.89 (1H, s), 7.29–7.33 (10H, m). EI MS m/z 676 (M^+). Anal. Calcd. for $\text{C}_{45}\text{H}_{72}\text{O}_4$ (677.1): C, 79.83; H, 10.72. Found: C, 79.52; H, 11.02.

(3R)-Diphenylmethyl 15-methyl-3-(13-methyltetradecanoyloxy)hexadecanoate (13R).

The same treatment mentioned above of **12R** (336 mg) gave **13R** (449 mg) in 89% yield. $[\alpha]_{\text{D}}^{24} -1.1^\circ$ (c 0.2, CHCl_3). MS m/z 676 (M^+). Anal. Calcd. for $\text{C}_{45}\text{H}_{72}\text{O}_4$ (677.1): C, 79.83; H, 10.72. Found: C, 79.53; H, 11.01.

(3S)-15-Methyl-3-(13-methyltetradecanoyloxy)hexadecanoic acid (14S).

A solution of **13S** (537 mg, 0.80 mmol) in EtOH (10 ml) containing 20% $\text{Pd}(\text{OH})_2$ on carbon as a catalyst (120 mg) was stirred under hydrogen atmosphere for 2 h at room temperature and filtered to give a crude mixture, which was chromatographed on a silica gel column. Elution with cyclohexane-EtOAc (10:1) gave **14S** (369 mg, 91%) as wax. The crude mixture contained diphenylmethane, but this did not affect in next reaction. Therefore, the mixture was employed for the next reaction without chromatographic purification. $[\alpha]_{\text{D}}^{24} +2.2^\circ$ (c 0.5, CHCl_3). IR ν_{max} (film) 3700–3000 (broad), 2926, 2855, 1740, 1713 cm^{-1} . 270 MHz ^1H NMR: (CDCl_3) δ 0.86 (12H, d, $J=6.6$ Hz), 1.11–1.63 (43H, m), 2.25–2.31 (2H, m), 2.58–2.62 (2H, m), 3.99 (1H, s), 5.21 (1H, m). Anal. Calcd. for $\text{C}_{32}\text{H}_{62}\text{O}_4$ (510.8): C, 75.24; H, 12.23. Found: C, 74.60; H, 12.07.

(3R)-15-Methyl-3-(13-methyltetradecanoyloxy)hexadecanoic acid (14R).

The same treatment as described above of **13R** gave **14R** in 90% yield as a wax. $[\alpha]_{\text{D}}^{24} -0.7^\circ$ (c 1.0, CHCl_3). Anal. Calcd. for $\text{C}_{32}\text{H}_{62}\text{O}_4$ (510.8): C, 75.24; H, 12.23. Found: C, 75.01; H, 12.29.

N-[(3S)-15-Methyl-3-(13-methyltetradecanoyloxy)hexadecanoyl]-L-serine benzyl ester (15SL). To a solution of **14S** (200 mg, 0.392 mmol) in CH_2Cl_2 (2 ml) was added oxalyl chloride (250 mg). After 1 h stirring at room temperature, the reaction mixture was concentrated *in vacuo* to give an acid chloride, which was dissolved in CH_2Cl_2 (4 ml). To this solution, L-serine benzyl ester hydrochloride (137 mg, 0.591 mmol) and Et_3N (100 mg, 0.990 mmol) were added under nitrogen and stirred for 1 h at 0–5 $^\circ\text{C}$. The reaction mixture was concentrated *in vacuo* and dissolved in EtOAc. The solution was washed with brine, dried over MgSO_4 , filtered, and concentrated *in vacuo* to give a mixture, which was chromatographed on a silica gel column. Elution with cyclohexane-EtOAc (3:1) gave **15SL** (174 mg, 65%) as a wax. $[\alpha]_{\text{D}}^{24} +10.5^\circ$ (c 0.23, CHCl_3). IR ν_{max} (film) cm^{-1} . 270 MHz ^1H NMR: (CDCl_3) δ 0.86 (12H, d, $J=6.6$ Hz), 1.11–1.72 (45H, m), 2.25–2.33 (2H, m), 2.48–2.52 (2H, m), 3.94 (2H, d, $J=3.3$ Hz), 4.64 (1H, td, $J=3.3, 7.1$ Hz, changed to a triplet on addition of D_2O), 5.21 (1H, m), 5.23 (2H, s), 6.64 (1H, d, $J=7.1$ Hz, NH), 7.36 (5H, bs). FAB MS (positive) m/z 688 [$\text{M}+\text{H}$] $^+$. High Resolution FAB MS (positive) m/z : Calcd. for $\text{C}_{42}\text{H}_{74}\text{NO}_6$, 688.5516; Found, 688.5513. Anal. Calcd. for $\text{C}_{42}\text{H}_{73}\text{NO}_6$ (688.0): C, 73.32; H, 10.70; N, 2.04. Found: C, 73.03; H, 10.65; N, 2.04.

***N*-[(3*S*)-15-Methyl-3-(13-methyltetradecanoyloxy)hexadecanoyl]-D-serine benzyl ester (15SD).** The same treatment as described above of **14S** with D-serine benzyl ester hydrochloride gave **15SD**. $[\alpha]_{\text{D}}^{24}$ -9.8° (c 0.23, CHCl₃). FAB MS (positive) m/z 688 [M+H]⁺. High Resolution FAB MS (positive) m/z : Calcd. for C₄₂H₇₄NO₆, 688.5516; Found, 688.5562. Anal. Calcd. for C₄₂H₇₃NO₆ (688.0): C, 73.32; H, 10.70; N, 2.04. Found: C, 73.44; H, 10.74; N, 2.09.

***N*-[(3*R*)-15-Methyl-3-(13-methyltetradecanoyloxy)hexadecanoyl]-L-serine benzyl ester (15RL).** The same treatment as described above of **14R** with L-serine benzyl ester hydrochloride gave **15RL** in 62% yield as a wax. $[\alpha]_{\text{D}}^{24}$ $+10.0^{\circ}$ (c 0.19, CHCl₃). 270 MHz ¹H NMR: (CDCl₃) δ 0.86 (12H, d, $J=6.6$ Hz), 1.11-1.70 (45H, m), 2.27-2.33 (2H, m), 2.50 (2H, d, $J=5.8$ Hz), 3.91-4.04 (2H, broad), 4.66 (1H, td, $J=3.4, 6.8$ Hz, changed to a triplet on addition of D₂O), 5.17 (1H, quintet, $J=6.2$ Hz), 5.22 (2H, s), 6.57 (1H, d, $J=7.0$ Hz, NH), 7.36 (5H, bs). FAB MS (positive) m/z 688 [M+H]⁺. High Resolution FAB MS (positive) m/z : Calcd. for C₄₂H₇₄NO₆, 688.5516; Found, 688.5516. Anal. Calcd. for C₄₂H₇₃NO₆ (688.0): C, 73.32; H, 10.70; N, 2.04. Found: C, 73.22; H, 10.73; N, 2.07.

***N*-[(3*S*)-15-Methyl-3-(13-methyltetradecanoyloxy)hexadecanoyl]-D-serine benzyl ester (15RD).** The same treatment as described above of **14R** with D-serine benzyl ester hydrochloride gave **15RD** as a wax. $[\alpha]_{\text{D}}^{24}$ -12.5° (c 0.24, CHCl₃). FAB MS (positive) m/z 688 [M+H]⁺. High Resolution FAB MS (positive) m/z : Calcd. for C₄₂H₇₄NO₆, 688.5516; Found, 688.5580. Anal. Calcd. for C₄₂H₇₃NO₆ (688.0): C, 73.32; H, 10.70; N, 2.04. Found: C, 73.61; H, 10.75; N, 2.04.

***N*-[(3*S*)-15-Methyl-3-(13-methyltetradecanoyloxy)hexadecanoyl]-L-serine (16SL).** A solution of **15SL** (156 mg, 0.227 mmol) containing 10% Pd on carbon (20 mg) as a catalyst in EtOAc (3 ml) was stirred under hydrogen atmosphere at room temperature for 2 h. After filtration, the solution was concentrated *in vacuo* to give **16SL** (135 mg, 99%) as a wax. $[\alpha]_{\text{D}}^{24}$ $+12.8^{\circ}$ (c 0.1, CHCl₃). 270 MHz ¹H NMR: (CDCl₃) δ 0.86 (12H, d, $J=6.6$ Hz), 1.12-1.66 (44H, m), 2.31 (2H, t, $J=7.5$ Hz), 2.48-2.63 (2H, m), 3.25-3.28 (2H, broad, OH x 2), 3.88 (1H, dd, $J=2.9, 11.7$ Hz), 4.08 (1H, m), 4.55 (1H, m), 5.24 (1H, m), 6.89 (1H, d, $J=6.6$ Hz, NH). FAB MS (positive) m/z 620 [M+Na]⁺, 598 [M+H]⁺. High Resolution FAB MS (positive) m/z : Calcd. for C₃₅H₆₈NO₆, 598.5047; Found, 598.5057. Anal. Calcd. for C₃₅H₆₇NO₆ (597.9): C, 70.31; H, 11.30; N, 2.34. Found: C, 70.37; H, 11.27; N, 2.38.

***N*-[(3*S*)-15-Methyl-3-(13-methyltetradecanoyloxy)hexadecanoyl]-D-serine (16SD).**

Compound **15SD** was hydrogenolyzed as described above to give **16SD** in 97% yield as a wax. $[\alpha]_{\text{D}}^{24}$ -13.7° (c 0.18, CHCl₃). 270 MHz ¹H NMR: (CDCl₃) δ 0.86 (12H, d, $J=6.6$ Hz), 1.12-1.63 (44H, m), 2.31 (2H, t, $J=7.5$ Hz), 2.55 (2H, d, $J=5.6$ Hz), 2.59-2.70 (2H, broad, OHx2), 3.88 (1H, d, $J=10.1$ Hz), 4.10 (1H, d, $J=10.4$ Hz), 4.56 (1H, s), 5.18 (1H, t, $J=6.1$ Hz), 6.83 (1H, d, $J=5.8$ Hz, NH). FAB MS (positive) m/z 620 [M+Na]⁺, 598 [M+H]⁺. High Resolution FAB MS (positive) m/z : Calcd. for C₃₅H₆₈NO₆, 598.5047; Found, 598.5069. Anal. Calcd. for C₃₅H₆₇NO₆ (597.9): C, 70.31; H, 11.30; N, 2.34. Found: C, 70.57; H, 11.23; N, 2.41.

***N*-[(3*R*)-15-Methyl-3-(13-methyltetradecanoyloxy)hexadecanoyl]-L-serine (16RL).**

Compound **15RL** was hydrogenolyzed as described above to give **16RL** in 95% yield as a wax. $[\alpha]_{\text{D}}^{24}$ $+13.3^{\circ}$ (c 0.14, CHCl₃). ¹H NMR spectra of **16SD** and **16RL** were identical. FAB MS (positive) m/z 620 [M+Na]⁺, 598 [M+H]⁺. High Resolution FAB MS (positive) m/z : Calcd. for C₃₅H₆₈NO₆, 598.5047; Found, 598.5071. Anal. Calcd. for C₃₅H₆₇NO₆ (597.9): C, 70.31; H, 11.30; N, 2.34. Found: C, 70.05; H, 11.37; N, 2.27.

***N*-[*N*-[(3*R*)-15-Methyl-3-(13-methyltetradecanoyloxy)hexadecanoyl]-D-serine (16*RD*).**

Compound **15*RD*** was hydrogenolyzed as described above to give **16*RD*** in 99% yield as a wax. $[\alpha]_{\text{D}}^{24} -14.4^{\circ}$ (c 0.18, CHCl₃). ¹H NMR spectra of **16*RD*** and **16*SL*** were identical. FAB MS (positive) *m/z* 620 [M+Na]⁺, 598 [M+H]⁺. High Resolution FAB MS (positive) *m/z*: Calcd. for C₃₅H₆₈NO₆, 598.5047; Found, 598.5071. *Anal.* Calcd. for C₃₅H₆₇NO₆ (597.9): C, 70.31; H, 11.30; N, 2.34. Found: C, 70.25; H, 11.33; N, 2.30.

N*-[*N*-[(3*S*)-15-Methyl-3-(13-methyltetradecanoyloxy)hexadecanoyl]-L-seryl]glycine benzyl ester (17*SL*).** To a solution of **16*SL (56 mg, 0.094 mmol) and glycine benzyl ester (31 mg, 0.188 mmol) in CH₂Cl₂ (30 ml) was added DCC (39 mg, 0.188 mmol) under nitrogen. The mixture was stirred for 16 h at room temperature, concentrated *in vacuo*, and diluted with EtOAc. The solution was washed with aq. NaHCO₃ and brine, dried over MgSO₄, filtered, and concentrated *in vacuo* to give a mixture, which was chromatographed on a silica gel column. Elution with benzene-EtOAc (1:1) gave **17*SL*** (63 mg, 90%) as a wax. IR ν_{max} (KBr) 3303, 2954, 2923, 2852, 1744, 1727, 1641 cm⁻¹. 270 MHz ¹H NMR: (CDCl₃) δ 0.86 (12H, d, *J*=6.6 Hz), 1.12-1.17 (4H, m), 1.25 (34H, s), 1.51 (2H, septet, *J*=6.6 Hz), 1.55-1.68 (4H, m), 2.26-2.31 (2H, m), 2.43-2.51 (2H, m), 3.11-3.21 (1H, broad m, OH), 3.61 (1H, m), 3.98-4.16 (3H, m), 4.47 (1H, m), 5.18 (2H, s), 5.20 (1H, m), 6.71 (1H, d, *J*=7.2 Hz, NH), 7.18 (1H, t, *J*=5.7 Hz, NH), 7.32-7.40 (5H, m). FAB MS (positive) *m/z* 767 [M+Na]⁺, 745 [M+H]⁺. High Resolution FAB MS (positive) *m/z*: Calcd. for C₄₄H₇₆N₂O₇Na, 767.5550; Found, 767.5552. *Anal.* Calcd. for C₄₄H₇₆N₂O₇ (745.1): C, 70.91; H, 10.29; N, 3.76. Found: C, 70.73; H, 10.46; N, 3.78.

N*-[*N*-[(3*S*)-15-Methyl-3-(13-methyltetradecanoyloxy)hexadecanoyl]-D-seryl]glycine benzyl ester (17*SD*).** Compound **16*SD was treated as described above to give **17*SD*** in 70% yield as a wax. $[\alpha]_{\text{D}}^{24} +12.4^{\circ}$ (c 0.50, CHCl₃). IR ν_{max} (KBr) 3299, 2954, 2922, 2852, 1727, 1640 cm⁻¹. 270 MHz ¹H NMR: (CDCl₃) δ 0.86 (12H, d, *J*=6.6 Hz), 1.12-1.17 (4H, m), 1.25 (34H, s), 1.51 (2H, septet, *J*=6.6 Hz), 1.55-1.62 (4H, m), 2.26-2.31 (2H, m), 2.46-2.49 (2H, m), 3.17-3.23 (1H, broad, OH), 3.62 (1H, m), 3.99-4.15 (3H, m), 4.50 (1H, m), 5.17-5.22 (3H, m), 6.72 (1H, d, *J*=7.3 Hz, NH), 7.25 (1H, t, *J*=5.7 Hz, NH), 7.32-7.40 (5H, m). FAB MS (positive) *m/z* 767 [M+Na]⁺, 745 [M+H]⁺. High Resolution FAB MS (positive) *m/z*: Calcd. for C₄₄H₇₇N₂O₇, 745.5731; Found, 745.5716. *Anal.* Calcd. for C₄₄H₇₆N₂O₇ (745.1): C, 70.91; H, 10.29; N, 3.76. Found: C, 71.14; H, 10.44; N, 3.76.

N*-[*N*-[(3*R*)-15-Methyl-3-(13-methyltetradecanoyloxy)hexadecanoyl]-L-seryl]glycine benzyl ester (17*RL*).** Compound **16*RL was treated as described above to give **17*RL*** in 70% yield as a wax. $[\alpha]_{\text{D}}^{24} -16.4^{\circ}$ (c 0.50, CHCl₃). IR and ¹H NMR spectra of **17*RL*** and **17*SD*** were identical. FAB MS (positive) *m/z* 767 [M+Na]⁺, 745 [M+H]⁺. High Resolution FAB MS (positive) *m/z*: Calcd. for C₄₄H₇₇N₂O₇, 745.5731; Found, 745.5711. *Anal.* Calcd. for C₄₄H₇₆N₂O₇ (745.1): C, 70.91; H, 10.29; N, 3.76. Found: C, 70.97; H, 10.39; N, 3.78.

N*-[*N*-[(3*R*)-15-Methyl-3-(13-methyltetradecanoyloxy)hexadecanoyl]-D-seryl]glycine benzyl ester (17*RD*).** Compound **16*RD was treated as described above to give **17*RD*** in 91% yield as a wax. IR and ¹H NMR spectra of **17*SL*** and **17*RD*** were identical. FAB MS (positive) *m/z* 767 [M+Na]⁺, 745 [M+H]⁺. High Resolution FAB MS (positive) *m/z*: Calcd. for C₄₄H₇₆N₂O₇Na, 767.5550; Found, 767.5510. *Anal.* Calcd. for C₄₄H₇₆N₂O₇ (745.1): C, 70.91; H, 10.29; N, 3.76. Found: C, 70.77; H, 10.30; N, 3.87.

N*-[*N*-[(3*S*)-15-Methyl-3-(13-methyltetradecanoyloxy)hexadecanoyl]-L-seryl]glycine (18*SL*).** A solution of **17*SL (16 mg, 0.021 mmol) containing 10% Pd on carbon (16 mg) as a catalyst in

EtOAc (3 ml) was stirred under hydrogen atmosphere at room temperature for 2.5 h. The reaction mixture was filtered through Celite and concentrated *in vacuo* to give **18SL** (10 mg, 71%) as a wax, which was a 3:1 mixture of amide rotational cis-trans isomers. $[\alpha]_{\text{D}}^{24} -8.0^{\circ}$ (c 0.23, CHCl_3). IR $\nu_{\text{max}}(\text{KBr})$ 3350, 3309, 2955, 2922, 2852, 1722, 1699, 1621 cm^{-1} . 400 MHz ^1H NMR: (CDCl_3) δ 0.86 (12H, d, $J=6.6$ Hz), 1.12-1.20 (4H, m), 1.25 (34H, bs), 1.51 (2H, septet, $J=6.6$ Hz), 1.56-1.65 (4H, m), 2.29 (2H, t, $J=7.1-7.8$ Hz), 2.52 (2H, d, $J=5.9$ Hz), 3.45 (2H, bs, OH and COOH), 3.68 (1H, dd, $J=4.6, 11.3$ Hz, serine CHOH), 3.96-4.10 (3H, m, glycine CH_2N and serine CHOH), 4.59 (1H, m, changed to a triplet on addition of D_2O , $J=4.2$ Hz, serine CH-N), 5.18 (1H, quintet, $J=5.8-6.8$ Hz, CH-OCO), 7.17 (3/4H, d, $J=7.4$ Hz, serine CONH), 7.22 (1/4H, d, $J=7.6$ Hz, serine CONH), 7.56 (3/4H, t, $J=5.3$ Hz, glycine CONH), 7.60 (1/4H, t, $J=5.3$ Hz, glycine CONH). FAB MS (positive) m/z 677 $[\text{M}+\text{Na}]^+$, 655 $[\text{M}+\text{H}]^+$. High Resolution FAB MS (positive) m/z : Calcd. for $\text{C}_{37}\text{H}_{71}\text{N}_2\text{O}_7$, 655.5261; Found, 655.5267.

***N*-[*N*-[(3*S*)-15-Methyl-3-(13-methyltetradecanoyloxy)hexadecanoyl]-D-seryl]glycine**

(**18SD**). Compound **17SD** was hydrogenolyzed as described above to give **18SD** in 84% yield as a wax, which was a 3:1 mixture of amide rotational cis-trans isomers. $[\alpha]_{\text{D}}^{24} +7.1^{\circ}$ (c 0.63, CHCl_3). IR $\nu_{\text{max}}(\text{KBr})$ 3354, 3056, 2954, 2921, 2851, 1738, 1723, 1664 cm^{-1} . 400 MHz ^1H NMR: (CDCl_3) δ 0.86 (12H, d, $J=6.6$ Hz), 1.12-1.20 (4H, m), 1.25 (34H, bs), 1.51 (2H, septet, $J=6.6$ Hz), 1.55-1.65 (4H, m), 2.26-2.32 (2H, m), 2.47-2.57 (2H, m), 3.63-3.70 (1H, m, CHOH), 3.94-4.11 (3H, m, CHOH, CH_2N), 4.58 (1H, m, changed to a triplet on addition of D_2O , $J=4.2$ Hz, serine CHN), 5.15-5.26 (1H, m, CHOCO), 7.20 (1/4H, d, $J=7.4$ Hz, serine CONH), 7.29 (3/4H, d, $J=7.7$ Hz, serine NH), 7.59 (1/4H, t, $J=5.3$ Hz, glycine CONH), 7.64 (3/4H, t, $J=5.4$ Hz, glycine NH). FAB MS (positive) m/z 677 $[\text{M}+\text{Na}]^+$, 655 $[\text{M}+\text{H}]^+$. High Resolution FAB MS (positive) m/z : Calcd. for $\text{C}_{37}\text{H}_{71}\text{N}_2\text{O}_7$, 655.5261; Found, 655.5254

***N*-[*N*-[(3*R*)-15-Methyl-3-(13-methyltetradecanoyloxy)hexadecanoyl]-L-seryl]glycine**

(**18RL**). Compound **17RL** was hydrogenolyzed as described above to give **18RL** in 84% yield as a wax, which was a 3:1 mixture of amide rotational cis-trans isomers. $[\alpha]_{\text{D}}^{24} -7.8^{\circ}$ (c 0.50, CHCl_3). IR and ^1H NMR were identical with those of **18SD**. FAB MS (positive) m/z 677 $[\text{M}+\text{Na}]^+$, 655 $[\text{M}+\text{H}]^+$. High Resolution FAB MS (positive) m/z : Calcd. for $\text{C}_{37}\text{H}_{71}\text{N}_2\text{O}_7$, 655.5261; Found, 655.5255. *Anal.* Calcd. for $\text{C}_{37}\text{H}_{70}\text{N}_2\text{O}_7$ (655.0): C, 67.85; H, 10.77; N, 4.28. Found: C, 67.47; H, 10.72; N, 4.27.

***N*-[*N*-[(3*R*)-15-Methyl-3-(13-methyltetradecanoyloxy)hexadecanoyl]-D-seryl]glycine**

(**18RD**). Compound **17RD** was hydrogenolyzed as described above to give **18RD** in 85% yield as a wax, which was a single amide isomer. $[\alpha]_{\text{D}}^{24} +7.6^{\circ}$ (c 0.75, CHCl_3). IR $\nu_{\text{max}}(\text{KBr})$ 3350, 3308, 2955, 2922, 2852, 1735 (shoulder), 1721, 1699, 1621, 1576 cm^{-1} . 400 MHz ^1H NMR: (CDCl_3) δ 0.86 (12H, d, $J=6.6$ Hz), 1.12-1.20 (4H, m), 1.25 (34H, bs), 1.51 (2H, septet, $J=6.6$ Hz), 1.56-1.65 (4H, m), 2.29 (2H, t, $J=7.1-7.8$ Hz), 2.52 (2H, d, $J=5.9$ Hz), 3.45 (2H, broad, OH, COOH), 3.68 (1H, dd, $J=4.6, 11.3$ Hz, CHOH), 3.96-4.10 (3H, m, CHOH, CH_2N), 4.55-4.59 (1H, m, $J=4.3$ Hz, serine CHN), 5.18 (1H, quintet, $J=5.8-6.8$ Hz, CH-OCO), 7.18 (1H, d, $J=7.4$ Hz, serine CONH), 7.56 (1H, d, $J=5.3$ Hz, glycine NH). FAB MS (positive) m/z 677 $[\text{M}+\text{Na}]^+$, 655 $[\text{M}+\text{H}]^+$. High Resolution FAB MS (positive) m/z : Calcd. for $\text{C}_{37}\text{H}_{70}\text{N}_2\text{O}_7\text{Na}$, 677.5081; Found, 677.5073. *Anal.* Calcd. for $\text{C}_{37}\text{H}_{70}\text{N}_2\text{O}_7$ (655.0): C, 67.85; H, 10.77; N, 4.28. Found: C, 67.47; H, 10.84; N, 4.44.

***N*-[(3*S*)-15-Methyl-3-(13-methyltetradecanoyloxy)hexadecanoyl]glycine benzyl ester**

(**19S**). To a solution of **14S** (400 mg, 0.783 mmol) in CH_2Cl_2 (4 ml) was added oxalyl chloride (500 mg, 3.94 mmol). After 1 h stirring at room temperature, the reaction mixture was concentrated *in vacuo* to give an acid chloride, which was dissolved in CH_2Cl_2 (4 ml). To this solution, glycine benzyl ester hydrochloride

(236 mg, 1.175 mmol) and Et₃N (200 mg, 1.980 mmol) were added under nitrogen atmosphere and stirred for 1 h at 0–5 °C. The reaction mixture was concentrated *in vacuo* and dissolved in EtOAc. The solution was washed with brine, dried over MgSO₄, filtered, and concentrated *in vacuo* to give a mixture, which was chromatographed on a silica gel column. Elution with benzene-EtOAc (9:1) gave **19S** (294 mg, 57%) as a wax. IR ν_{\max} (film) 3310, 2954, 2921, 2852, 1737, 1729, 1640, 1549 cm⁻¹. 400 MHz ¹H NMR: (CDCl₃) δ 0.86 (12H, d, $J=6.6$ Hz), 1.12–1.17 (4H, m), 1.25 (34H, bs), 1.51 (2H, septet, $J=6.6$ Hz), 1.57–1.66 (4H, m), 2.30 (2H, t, $J=7.5$ –7.8 Hz), 2.47–2.57 (2H, m), 4.07 (2H, d, $J=5.1$ Hz), 5.16 (1H, m), 5.19 (2H, s), 6.26 (1H, t, $J=4.9$ Hz, NH), 7.34–7.40 (5H, m). FAB MS (positive) m/z 680 [M+Na]⁺, 658 [M+H]⁺. High Resolution FAB MS (positive) m/z : Calcd. for C₄₁H₇₂NO₅, 658.5410; Found, 658.5388. Anal. Calcd. for C₄₁H₇₁NO₅ (658.0): C, 74.84; H, 10.88; N, 2.13. Found: C, 74.72; H, 11.00; N, 2.24.

N-[(3R)-15-Methyl-3-(13-methyltetradecanoyloxy)hexadecanoyl]glycine benzyl ester (19R). The same reaction as described above of **14R** gave **19R** in 69% as a wax. $[\alpha]_{\text{D}}^{24} +0.2^\circ$ (c 0.50, CHCl₃). IR and ¹H NMR spectra of **19R** and **19S** were identical. FAB MS (positive) m/z 680 [M+Na]⁺, 658 [M+H]⁺. High Resolution FAB MS (positive) m/z : Calcd. for C₄₁H₇₁NO₅Na, 680.5230; Found, 680.5235. Anal. Calcd. for C₄₁H₇₁NO₅ (658.0): C, 74.84; H, 10.88; N, 2.13. Found: C, 74.73; H, 10.60; N, 2.31.

N-[(3S)-15-Methyl-3-(13-methyltetradecanoyloxy)hexadecanoyl]glycine (20S). A solution of **19S** (253 mg, 0.38 mmol) containing 10% Pd on carbon (30 mg) as a catalyst in EtOAc (6 ml) was stirred under hydrogen at room temperature for 2 h. The reaction mixture was filtered through Celite and concentrated *in vacuo* to give **20S** (205 mg, 94%) as a wax. IR ν_{\max} (film) 3363, 2955, 2919, 2850, 1724, 1627, 1549 cm⁻¹. 400 MHz ¹H NMR: (CDCl₃) δ 0.86 (12H, d, $J=6.6$ Hz), 1.12–1.17 (4H, m), 1.25 (34H, bs), 1.51 (2H, septet, $J=6.6$ Hz), 1.56–1.67 (4H, m), 2.31 (2H, t, $J=7.5$ –7.7 Hz), 2.49–2.59 (2H, m), 4.07 (2H, d, $J=5.2$ Hz), 5.16 (1H, quintet, $J=5.5$ –6.9 Hz, CHOCO), 6.41 (1H, t, $J=5.2$ Hz, NH). FAB MS (positive) m/z 590 [M+Na]⁺, 568 [M+H]⁺. High Resolution FAB MS (positive) m/z : Calcd. for C₃₄H₆₆NO₅, 568.4941; Found, 568.4958. Anal. Calcd. for C₃₄H₆₅NO₅ (567.9): C, 71.91; H, 11.54; N, 2.47. Found: C, 71.90; H, 11.66; N, 2.53.

N-[(3R)-15-Methyl-3-(13-methyltetradecanoyloxy)hexadecanoyl]glycine (20R). The same hydrogenolysis as described above of **19R** gave **20R** in 98% yield. IR and ¹H NMR spectra of **20R** and **20S** were identical. FAB MS (positive) m/z 590 [M+Na]⁺, 568 [M+H]⁺. High Resolution FAB MS (positive) m/z : Calcd. for C₃₄H₆₆NO₅, 568.4941; Found, 568.4944.

N-[N-[(3S)-15-Methyl-3-(13-methyltetradecanoyloxy)hexadecanoyl]glycyl]-L-serine benzyl ester (21SL). To a solution of **20S** (80 mg, 0.14 mmol) and L-serine benzyl ester (82 mg, 0.42 mmol) in CH₂Cl₂ (4 ml) was added 1,3-dicyclohexylcarbodiimide (87 mg, 0.42 mmol) under nitrogen. The mixture was stirred for 2 h at room temperature, concentrated *in vacuo*, and then diluted with EtOAc. The solution was washed with aq. NaHCO₃ and brine, dried over MgSO₄, filtered, and concentrated *in vacuo* to give a mixture, which was chromatographed on a silica gel column. Elution with benzene-EtOAc (1:1) gave **21SL** (80 mg, 76%) as a wax. $[\alpha]_{\text{D}}^{24} +25.0^\circ$ (c 0.5, CHCl₃). IR ν_{\max} (KBr) 3398, 3330, 2954, 2923, 2852, 1738, 1727, 1707(w), 1657, 1637, 1615 (w), 1538 cm⁻¹. 400 MHz ¹H NMR: (CDCl₃) δ 0.86 (12H, d, $J=6.6$ Hz), 1.12–1.17 (4H, m), 1.25 (34H, s), 1.51 (2H, septet, $J=6.6$ Hz), 1.55–1.63 (4H, m), 2.31 (2H, t, $J=7.5$ –7.8 Hz), 2.47 (2H, d, $J=6.2$ Hz), 3.35 (1H, dd, $J=5.8$, 7.4 Hz, OH), 3.88–4.12 (4H, m), 4.72 (1H, td, $J=3.6$, 7.8 Hz), 5.21 (1H, m, CHOCO), 5.22 (2H, s), 6.49 (1H, t, $J=5.5$ Hz, NH), 7.17 (1H, d, $J=7.7$ Hz, NH), 7.33–7.38 (5H, m). FAB MS (positive) m/z 767 [M+Na]⁺, 745 [M+H]⁺. High Resolution FAB MS (positive) m/z :

Calcd. for $C_{44}H_{77}N_2O_7$, 745.5731; Found, 745.5722. *Anal.* Calcd. for $C_{44}H_{76}N_2O_7$ (745.1): C, 70.93; H, 10.28; N, 3.76. Found: C, 70.63; H, 10.00; N, 3.81.

***N*-[*N*-[(3*S*)-15-Methyl-3-(13-methyltetradecanoyloxy)hexadecanoyl]glycyl]-D-serine benzyl ester (21SD).** The same procedure as described above of 20*S* and D-serine benzyl ester gave 21SD in 70% yield as a wax. $[\alpha]_D^{24}$ -10.6° (c 0.5, $CHCl_3$). IR $\nu_{max}(KBr)$ 3319, 2954, 2922, 2852, 1733, 1661, 1637, 1549 cm^{-1} . 400 MHz 1H NMR: ($CDCl_3$) δ 0.86 (12H, d, $J=6.6$ Hz), 1.12-1.17 (4H, m), 1.25 (34H, bs), 1.51 (2H, septet, $J=6.6$ Hz), 1.56-1.63 (4H, m), 2.30 (2H, t, $J=7.5$ Hz), 2.48 (2H, d, $J=5.9$ Hz), 3.18 (1H, t, $J=5.8$ Hz, OH), 3.94-4.05 (4H, m), 4.70 (1H, td, $J=3.4, 7.6$ Hz), 5.15 (1H, quintet, $J=5.8$ Hz, CHOCO), 5.21 (2H, s), 6.48 (1H, t, $J=5.4$ Hz, glycine NH), 7.05 (1H, d, $J=7.6$ Hz, serine NH), 7.33-7.39 (5H, m). FAB MS (positive) m/z 767 [M+Na]⁺, 745 [M+H]⁺. High Resolution FAB MS (positive) m/z : Calcd. for $C_{44}H_{77}N_2O_7$, 745.5731; Found, 745.5734. *Anal.* Calcd. for $C_{44}H_{76}N_2O_7$ (745.1): C, 70.93; H, 10.28; N, 3.76. Found: C, 70.79; H, 10.36; N, 3.80.

***N*-[*N*-[(3*R*)-15-Methyl-3-(13-methyltetradecanoyloxy)hexadecanoyl]glycyl]-L-serine benzyl ester (21RL).** The same procedure as described above of 20*R* and L-serine benzyl ester gave 21RL in 72% yield as a wax. $[\alpha]_D^{24}$ +10.2° (c 0.5, $CHCl_3$). IR and 1H NMR spectra of 21RL and 21SD were identical. FAB MS (positive) m/z 767 [M+Na]⁺, 745 [M+H]⁺. High Resolution FAB MS (positive) m/z : Calcd. for $C_{44}H_{77}N_2O_7$, 745.5731; Found, 745.5720. *Anal.* Calcd. for $C_{44}H_{76}N_2O_7$ (745.1): C, 70.93; H, 10.28; N, 3.76. Found: C, 71.02; H, 10.29; N, 3.81.

***N*-[*N*-[(3*R*)-15-Methyl-3-(13-methyltetradecanoyloxy)hexadecanoyl]glycyl]-D-serine benzyl ester (21RD).** The same procedure as described above of 20*R* and D-serine benzyl ester gave 21RD in 82% yield as a wax. $[\alpha]_D^{24}$ -24.0° (c 0.5, $CHCl_3$). IR and 1H NMR spectra of 21SL and 21RD were identical. FAB MS (positive) m/z 767 [M+Na]⁺, 745 [M+H]⁺. High Resolution FAB MS (positive) m/z : Calcd. for $C_{44}H_{77}N_2O_7$, 745.5731; Found, 745.5725. *Anal.* Calcd. for $C_{44}H_{76}N_2O_7$ (745.1): C, 70.93; H, 10.28; N, 3.76. Found: C, 71.27; H, 10.15; N, 3.81.

***N*-[*N*-[(3*S*)-15-Methyl-3-(13-methyltetradecanoyloxy)hexadecanoyl]glycyl]-L-serine (22SL).** A solution of 21SL (32 mg) in EtOAc (4 ml) containing 10% Pd on carbon (32 mg) was hydrogenolized under hydrogen for 2.5 h at room temperature. The solution was filtered through Celite and concentrated *in vacuo* to give 22SL (14 mg, 50%) as a wax. $[\alpha]_D^{24}$ +25.6° (c 0.38, $CHCl_3$). IR $\nu_{max}(KBr)$ 3371, 3335, 3280, 3073 (w), 2955, 2918, 2851, 1746, 1683, 1644 cm^{-1} . 400 MHz 1H NMR: ($CDCl_3$) δ 0.86 (12H, d, $J=6.6$ Hz), 1.12-1.20 (4H, m), 1.25 (34H, bs), 1.51 (2H, septet, $J=6.6$ Hz), 1.55-1.65 (4H, m), 2.30 (2H, t, $J=7.5$ Hz), 2.51 (2H, d, $J=5.5$ Hz), 3.88-4.12 (4H, m, CH_2OH, CH_2N), 4.61 (1H, broad s), 5.21 (1H, quintet, $J=5.8-6.2$ Hz, CHOCO), 5.3-5.8 (2H, broad, OH, COOH), 7.18 (1H, broad s, NH), 7.60 (1H, broad s, NH). FAB MS (positive) m/z 699 [M+2Na-H]⁺, 677 [M+Na]⁺, 655 [M+H]⁺. High Resolution FAB MS (positive) m/z : Calcd. for $C_{37}H_{70}N_2O_7Na$, 677.5081; Found, 677.5092.

***N*-[*N*-[(3*S*)-15-Methyl-3-(13-methyltetradecanoyloxy)hexadecanoyl]glycyl]-D-serine (22SD).** The same hydrogenolysis as described above of 21SD gave 22SD in 73% yield as a wax. $[\alpha]_D^{24}$ -18.8° (c 0.52, $CHCl_3$). IR $\nu_{max}(KBr)$ 3318, 3079 (w), 2955, 2922, 2852, 1723, 1681, 1659, 1629, 1545 cm^{-1} . 400 MHz 1H NMR: ($CDCl_3$) δ 0.86 (12H, d, $J=6.6$ Hz), 1.12-1.20 (4H, m), 1.25 (34H, bs), 1.51 (2H, septet, $J=6.6$ Hz), 1.55-1.65 (4H, m), 2.30 (2H, t, $J=7.5$ Hz), 2.46-2.57 (2H, m), 3.88-4.12 (4H, m, CH_2OH, CH_2N), 4.63 (1H, broad s, CHN), 5.18 (1H, quintet, $J=5.6-5.9$ Hz, CHOCO), 5.60 (2H, broad, OH, COOH), 7.16 (1H, broad s, glycine NH), 7.62 (1H, d, $J=6.9$ Hz, serine NH). FAB MS (positive) m/z

699 [M+2Na-H]⁺, 677 [M+Na]⁺, 655 [M+H]⁺. High Resolution FAB MS (positive) *m/z*: Calcd. for C₃₇H₇₁N₂O₇, 655.5261; Found, 655.5271.

***N*-[*N*-[(3*R*)-15-Methyl-3-(13-methyltetradecanoyloxy)hexadecanoyl]glycyl]-L-serine (22RL).** The same hydrogenolysis as described above of 21RL gave 22RL in 70% yield as a wax. [α]_D²⁴ +18.9° (c 0.39, CHCl₃). IR and ¹H NMR spectra of 22RL and 22SD were identical. And 500 MHz ¹H NMR and FAB MS data of 22RL and natural flavolipin were identical. FAB MS (positive) *m/z* 699 [M+2Na-H]⁺, 677 [M+Na]⁺, 655 [M+H]⁺. High Resolution FAB MS (positive) *m/z*: Calcd. for C₃₇H₇₀N₂O₇Na, 677.5081; Found, 677.5090. The macrophage stimulation activity of 22RL was almost the same as that of natural flavolipin, whereas 22SD was practically inactive.

***N*-[*N*-[(3*R*)-15-Methyl-3-(13-methyltetradecanoyloxy)hexadecanoyl]glycyl]-D-serine (22RD).** The same hydrogenolysis as described above of 21RD gave 22RD in 47% yield as a wax. [α]_D²⁴ -26.2° (c 0.58, CHCl₃). IR and ¹H NMR spectra of 22SL and 22RD were identical. FAB MS (positive) *m/z* 699 [M+2Na-H]⁺, 677 [M+Na]⁺, 655 [M+H]⁺. High Resolution FAB MS (positive) *m/z*: Calcd. for C₃₇H₇₁N₂O₇, 655.5261; Found, 655.5266.

Acknowledgement. This study was supported, in part, by the Social Insurance Agency Fund commissioned by Japan Health Sciences Foundation.

References and Notes

- [1] Kawai, Y.; Yano, I.; Kaneda, K. *Eur. J. Biochem.* **1988**;171:73-80.
- [2] Kawai, Y.; Akagawa, K. *Infection and Immunity.* **1989**;57:2086-2091.
- [3] Kawai, Y.; Kamoshita, K.; Akagawa, K. *FEMS Microbiology Lett.* **1991**;83:127-130.
- [4] Kawai, Y.; Kaneda, K.; Morisawa, Y.; Akagawa, K. *Infection and Immunity.* **1991**;59:2560-2566.
- [5] Shiozaki, M.; Deguchi, N.; Mochizuki, T.; Nishijima, M. *Tetrahedron Lett.* **1996**;37:3875-3876.
- [6] Shiozaki, M.; Deguchi, N.; Ishikawa, T.; Haruyama, H.; Kawai, Y.; Nishijima, M. *Tetrahedron Lett.* **1998**;39:4497-4500.
- [7] Uchida, I.; Yoshida, K.; Kawai, Y.; Takasa, S.; Itoh, Y.; Tanaka, H.; Kohsaka, M.; Imanaka, H. *J. Antibiotics.* **1985**;38:1476-1486.
- [8] Nemoto, T.; Ojika, M.; Takahata, Y.; Andoh, T.; Sakagami, Y. *Tetrahedron.* **1998**;54:2683-2690.
- [9] Freudenberg, K.; Wolf, A. *Ber.* **1927**;60:232-238.
- [10] Barton, DHR.; McCombie, SW. *J. Chem. Soc. Perkin Trans I.* **1975**:1574-1585.
- [11] Murray, DH.; Prokop, J. *J. Pharm. Sci.* **1965**;54:1468-1473.
- [12] Uchida, I.; Yoshida, K.; Kawai, Y.; Takase, S.; Itoh, Y.; Tanaka, H.; Kosaka, M.; Imanaka, H. *Chem. Pharm. Bull.* **1985**;33:424-427.
- [13] Compound (20S) has been reported as belonging to the N-type calcium channel blockers from a marine bacterium, *Cytophaga* sp. SANK 71996. Morishita, T.; Sato, A.; Hisamoto, M.; Oda, M.; Matsuda, K.; Ishii, A.; Kodama, K. *J. Antibiotics.* **1997**;50:457- 468.
- [14] Compound (20R) was already reported in reference [7].
- [15] Yoshida, K.; Iwami, M.; Umehara, Y.; Nishikawa, M.; Uchida, I.; Kohsaka, M.; Aoki, H.; Imanaka, H. *J. Antibiotics.* **1985**;38:1469-1475.